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## Plant Tissue Culture: A Review

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### ABSTRACT

Plant tissue culture refers to growing and multiplication of cells, tissues and organs of plants on defined solid or liquid media under aseptic and controlled environment. The commercial technology is primarily based on micropropagation, in which rapid proliferation is achieved from tinystem cuttings, axillary buds, and to a limited extent from somatic embryos, cell clumps in suspension cultures and bioreactors. The cultured cells and tissue can take several pathways. The pathways that lead to the production of true-to-type plants in large numbers are the preferred ones for commercial multiplication. The process of micropropagation is usually divided into several stages i.e., prepropagation, initiation of explants, subculture of explants for proliferation, shooting and rooting, and hardening. These stages are universally applicable in large-scale multiplication of plants. The delivery of hardened small micropropagated plants to growers and market also requires extra care.

**Keywords:** multiplication of cells, micropropagation, pathways



### INTRODUCTION

A whole plant can be regenerated from a small tissue or plant cells in a suitable culture medium under controlled environment. The plantlets so produced are called tissue-culture raised plants. These plantlets are a true copy of the mother plant and show characteristics identical to the mother plant. For example, if the mother plant is a high yielding plant the plantlets will also be high yielding. Many plant species are presently being propagated through tissue culture successfully. This capacity of a single cell to grow into a complete plant is termed as Totipotency, which was first put forward by a German Botanist Haberlandt in 1902. Tissue culture is the propagation of plants wherein a part/tissue of the plant is placed in nutrient media that favors the production of shoots, roots following which they are hardened and transferred to soil. Quality planting material of economically important species can be produced in a large scale/desired quantity through tissue culture<sup>[1]</sup>. Plant tissue culture can be initiated from almost any part of a plant however, for micropropagation or direct shoot regeneration, meristematic tissue such as shoot tip is ideal. The physiological state of the plant does have an influence on its response to tissue culture. The mother plant must be healthy and free from

obvious signs of disease or pest. The shoot tip explants being juvenile contain a higher proportion of actively dividing cells. It is important to use quality mother plant stock to initiate cultures<sup>[2]</sup>. The cultural conditions required to initiate and sustain plant cells in culture, or to regenerate intact plants from cultured cells, are different for each plant species. Each variety or clone of a species often have a particular set of cultural requirements.

### REVIEW OF LITERATURE

Kato et al suggested that an agitation speed of 50 to 100 r.p.m. was most appropriate for the growth of tobacco cells in stirred-jar fermenters. It is true that culture plant cells are more fragile than bacterial cells, however, Martin noted: "it seems obvious that cells lines differ in their resistance to shear effect and that a single optimum agitation speed that cannot be designed for all lines"<sup>[3]</sup>. A roller-bottled system using a round flask was used by Lampert in 1964. A V-shape fermenter was proposed by Veliky and Martin. It is an inverted flask carrying two Teflon-coated stirring bars on a glass pin situated at the bottom of the flask. A drain/sample port is also located at the bottom. The top of the flask is fitted with four standard taper penetrations. Berlin et al compared the highest cinnamoyl putrescine producing, p-fluorophenylamine resistance

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strain TX-4 or *N. tabacum* L. CV xanthi with a low producing strain for five enzymes of the biosynthetic pathway. As a result, activities of these enzymes, phenylalanine ammonia-lyase, trans-cinnamate 4-hydroxylase, 4-coumarate:CoA ligase, ornithine decarboxylase and arginine decarboxylase were found to be 3 to 10 times higher in TX4 cells<sup>[4]</sup>.

Davis *et al* recognised that addition of oxalate to the medium of *Gossypium hirsutum* suspension culture could reduce the amount of verticillium dahlia elicitor to be employed to stimulate metabolite synthesis. Addition of a fungal elicitor often inhibits the growth of plant cells but a combination of the elicitor and oxalate did not reduce the cell mass of the plant, therefore secondary metabolite synthesis was increased up to ten fold<sup>[5]</sup>. Dunlop and Curtis reported that a combination of phosphate limitation and fungal elicitation synergistically produced secondary metabolites. They found that either phosphate limitation or elicitation with a mycelial extract of the fungus, *Rhizoctonia solani* alone results in increased production of the sesquiterpene solavetivone by *Agrobacterium rhizogenes*-transformed hairy root cultures of *Hyoscyamus muticus*. In many species, somatic embryos are morphologically similar to the zygotic embryos, although some biochemical, physiological and anatomical differences have been documented. The synthetic auxin, 2,4-D is commonly used for embryo induction in many angiosperms, e.g., carrot and alfalfa subculture of cells from 2,4-D containing medium to auxin-free medium is sufficient to induce somatic embryogenesis.

#### Stages of Tissue Culture Process:

**1. Preparation of nutrient medium:** A semi-solid medium is prepared in double distilled water containing macroelements, micro elements, amino acids, vitamins, iron source, carbon source like sucrose and phyto-hormones. The medium is heated for dissolving the agar and 25 to 50 ml is dispensed into each wide mouth bottle. The vessels containing culture media are then sealed and sterilized by autoclaving<sup>[6]</sup>.

**2. Establishment of aseptic culture:** The starting material for the process is normally an actively growing shoot tip of axillary or terminal bud or shoot tip of a plant. The process of tissue culture starts from the selection of mother plants having the desired characteristics. Ex-plant preferably the meristematic tissue of the selected mother plant is isolated. The excised tissue/explant is washed with water and then rinsed with a disinfectant such as Savlon or detol solution followed by a sterile-water wash. The tissue is then dipped in 10% bleach solution for ten minutes for disinfecting the plant tissue material, killing

most of the fungal and bacterial organisms. Sterilization process of explants depends on the plant species and types of explants<sup>[7]</sup>.

**3. Inoculation:** Inoculation is carried out under aseptic conditions. In this process explants or micro shoots are transferred to the sterilized nutrient medium.

**4. Development of plants in growth room:** After the inoculation of the plant tissue, the bottles are sealed and transferred into growth room to trigger developmental process under diffused light (fluorescent light of 1000-2000 lux) at  $25 \pm 2$  °C and 50 to 60% relative humidity. Light and temperature requirements vary from species to species and sometimes during the various stages of development. The cultures are observed daily for growth and any signs of infection/contamination. Cultures, that do not show good growth or infected, are discarded. The healthy cultures grow into small shoot buds. These are sub-cultured on the fresh medium after 4 weeks. The number of subcultures required is specific to the plant species, which are standardized. The shoots generally develop after 4 weeks. After enough number of shoots is developed in each container (10 to 15), to a minimum height of 2 cm they are transferred to another medium for initiating the process of rooting. The constituent of rooting medium for each plant species are specific. Roots are generally formed within 2 to 4 weeks. Plants at this stage are delicate and require careful handling<sup>[8]</sup>.

**5. Hardening of micro plants:** Due to very high humidity inside the culture vessel and artificial conditions of development, the plantlets are tender and are therefore not ready for coping up with the field conditions. The plants removed from the sterile medium are washed and are maintained under intermittent mist or are covered with clean transparent plastic. After 10 to 15 days under high humidity, the plants are transferred to green house and maintained for another 4 to 6 weeks. They are then ready to be transferred to net house or the field. Normally, the tissue culture plants are sold either as ex-agar plants or hardened plants from the green house.

**A: Ex-agar plants:** Depending on the parameters such as location/the site of planting, soil quality and the climatic conditions defined by the customer, the ex-agar plant for sale could be in vitro rooted plants or only the shoots. When the tissue culture plants are sold at this stage, the plants are washed in sterilized water to remove the agar medium. The washed plants are sorted into 2 to 3 grades and packed in corrugated plastic boxes lined with sterilized tissue paper as per specifications of the Plant Quarantine Authority, Government of India for exports. The number of plants per box depends on the customer's requirement. Depending on the final destination and the preference of the

customer, the plants are treated with specific fungicides and antibiotics to avoid infection. The ex-agar plants are preferred for export or for destinations where hardening facilities are available. The plants after being removed from nutrient media should preferably be transplanted within 72 hours<sup>[9]</sup>.

**B: Hardened plants:** The plants are transferred to net pots/ pro tray for acclimatization after they fully develop shoots and roots in the bottles. The rooted plantlets are transferred to pots filled with suitable substrate and are watered. This operation is carried out on an open bench. These pots are then transferred to the green house for 4 to 6 weeks. During this process, they are given fertilizers and treated like plantlets obtained by any other means of propagation. After the plants are acclimatized fully, they are transferred to poly-bags. At this stage the plants are completely hardened and are ready to be planted in the field for cultivation. Hardening units can be set up in sites away from the micropropagation unit<sup>[10]</sup>.

**5. Advantages of Micro-propagation Technology**  
Micro-propagation has several advantages over conventional methods of propagation such as:

**1. Rapid multiplication:** Micro-propagation offers rapid multiplication of desired plant species.

**2. Requirement of only limited number of explants:** Small pieces of plant (explants)/tissue can be used to produce a large number of plants in a relatively small space.

**3. Uniform or true to type plants:** Micro-propagation provides a high degree of phenotypic/physical uniformity. Since the production cycle takes place under controlled conditions, proper planning and scheduling based on the market demand is possible. The resulting product has very high degree of uniformity compared with traditionally propagated plants.

**4. Germplasm storage:** Plants can be stored *in vitro* in a small space and less labour is required for maintenance of stock plants.

**5. Disease free planting material:** Plantlets produced by tissue culture are usually disease free. With proper diagnosis and treatments, elimination of fungus, bacteria and virus prior to large scale propagation is possible. With the help of serological and molecular techniques it is possible to index virus of mother plant/explant which is to be used for mass multiplication.

**6. Growth manipulation:** Nutrient levels, light, temperature and other factors can be more effectively controlled to manipulate the growth, multiplication and regeneration.

**7. Round the year production:** Micro-propagation is independent of season. As micro-propagation could be carried out throughout the year; production cycle can be scheduled to meet peak demands. For species that have long generation time, low levels

of seed production, or seeds that do not readily germinate, rapid propagation is possible through tissue culture. The time required is much shortened, no need to wait for the whole life cycle of seed development. Commercially propagated plants through micro-propagation in India<sup>[11]</sup>. The plants in each category which are commercially propagated are as follows

Table 1. Type of plants.

Plant type	Name of plant
Medicinal plants	Aloe vera, Geranium, Stevia, Patchouli, Neem
Ornamentals	Gerbera, Carnation, Anthurium, Lily, Syngonium, Cymbidium
Woody Plants	Teak, Bamboo, Eucalyptus, Populus
Bio fuel	Jatropha, Pongamia

## 6. Mitigating Risks of commercial plant tissue culture:

The utilization of plant tissue culture for commercial production is limited by two major risks viz., spread of diseases especially those caused by viruses, and variations. The movement of plants also involves accidental risk of introducing plant disease. Pathogens that are often symptomless, such as viruses, pose a risk. The risk of distribution of inferior micropropagated plants has posed a major threat to the ever-increasing agribusiness industry. In order to prevent these risks, effective testing (indexing) procedures are required prior to bulking up culture for commercial propagation. Standard procedure should be adopted such as:

- Careful selection of mother plants
- Ensuring establishment of virus free culture through indexing of 100 % explants
- Proper package and practices to be adopted such as limited number of cycles of multiplication, grading of cultures as well as plants, insect, pest monitoring in hardening area etc<sup>[12]</sup>.

## 7. Need for Certification of tissue culture raised plants:

Micropropagation is effectively used for producing quality planting material free from disease. Yet there is threat of inadvertent propagation of virus infected plants which will not only result in loss or poor performance of the crop but also spread of virus. Further failure to use standard crop specific guidelines can lead to variations in the plants produced. The most deleterious variants in tissue culture raised plants are those that affect yield through somaclonal variations and carry viruses and other pathogens which are difficult to diagnose. This is an area of great concern and requires a well-structured system to support the tissue culture industry to ensure virus free quality planting material for commercial production. With the objective of production and

distribution of quality tissue culture planting materials Department of Biotechnology (DBT), Government of India has established National Certification System for Tissue Culture Raised Plants (NCS TCP). For details about NCS-TCP, please refer the manual on "National Certification System for Tissue Culture Raised Plants (NCS-TCP): An Overview or log in to www.dbtncstcp.nic.in . DBT is the Certification agency for the purpose for certification of Tissue culture raised plants/propagules up to laboratory level and to regulate its genetic fidelity as authorized vide the Gazette of India Notification dated 10 th March 2006 of Ministry of Agriculture under section 8 of the Seeds Act.

## **PART B: TECHNO-COMMERCIAL FEASIBILITY:**

**1. MARKET SCENARIO:** Demand for tissue cultured plantlets is growing rapidly. India, with its low cost skilled labour as well as scientific manpower (both of which are essential for tissue culture) has a natural advantage. Additional favourable factors are the wide range of plant biodiversity in the country and favorable tropical climate (which enables greenhouses with low energy consumption) The potential for the domestic market is enormous and by conservative estimates it is around Rs. 200 crores with an annual growth rate of 20%. There are more than 70 established commercial tissue culture units. Their production capacity ranges between 0.5 million to 10 million plants per annum with an aggregate production capacity of about 200 million plantlets per year. The protocols have either been developed in-house or transferred through the various research institutions and universities engaged in development of the protocols through support of the Department of Biotechnology (DBT) Currently, the focus of the companies is mainly banana, floriculture, sugarcane and potato. With increasing awareness about the advantages of tissue culture raised plants in improving yield and quality, their domestic consumption is also increasing optimistically. The major consumers of tissue culture raised plants are the State Agriculture Department, Agri Export Zones (AEZs), State agencies such as Spice Board, sugar industry and private farmers. The paper industry, medicinal plant industry and State Forest Departments are using tissue culture raised plants in a limited scale. Also a number of progressive farmers and nurseries in the states are the major consumers of Tissue culture plants particularly for flowers, banana, sugarcane and medicinal plants<sup>[13]</sup>.

## **2. Establishment of Commercial Plant Tissue Culture Unit**

Commercial plant tissue culture unit consists of the following components

**Storage room for chemicals:** It is advisable to have a separate area for storage of chemicals, apparatus and equipments. Chemicals required in small amounts should not be purchased in large quantities as they may lose their activity, pick up moisture or get contaminated. Such problems can be overcome by purchasing small lots on a regular basis. Washing and Media Preparation Room: The glassware washing area should be located near the sterilization room. This area should have at least one large sink but two sinks are preferable with running tap water. Adequate workspace is required on each side of the sink; this space is used for glassware soaking and drainage. Plastic netting can be placed on surfaces near the sink to reduce glassware breakage and enhance water drainage. The outlet pipe from the sink should be of PVC to resist damage from acids and alkalis. Both hot and cold water should be available and the water still and de-ionisation unit should be located nearby. The washing room should be swapped periodically. Mobile drying racks can be used and lined with cheesecloth to prevent water dripping and loss of small objects. Ovens or hot air cabinets should be located close to the glassware washing and storage area. Dust-proof cabinets and storage containers should be installed to allow for easy access to glassware. When culture vessels are removed from the growth area, they are often autoclaved to kill contaminants and to soften semi-solid media. It should be possible to move the vessels easily to the washing area. The glassware storage area should be close to the wash area to expedite storage and access for media preparation. The media preparation room should have smooth walls and floors, which enable easy cleaning to maintain a high degree of cleanliness. Minimum number of doors and windows should be provided in this room but within the local fire safety regulations. Media preparation area should be equipped with both tap and purified water. An appropriate system for water purification must be selected and fitted after careful consideration of the cost and quality. A number of electrical appliances are required for media preparation; hence, it is essential to have safety devices like fire extinguisher, fire blanket and a first aid kit in the media preparation room. A variety of glassware, plastic ware and stainless steel apparatus is required for measuring, mixing, and media storage. These should be stored in the cabinets built under the worktables and taken out for use as and when required. The water source and glassware storage area should be in or near the media preparation area<sup>[14]</sup>. The workbench tops should be made with plastic laminated surfaces that can tolerate frequent cleaning. Media storage room should have capacity to store the media for at least 7 days. Sterility Class 1,00,000 is desirable for

media storage room. Inoculation Room The most important work area is the Inoculation room where the core activity takes place. The transfer area needs to be as clean as possible with minimal air disturbance. Walls and floors of the Inoculation room must be smooth to ensure frequent cleaning.

**Profile of a self contained unit:** The project profile of a micropropagation unit with an annual production capacity of 3 million plantlets is discussed below. A product mix of 5 different plants has been assumed:

1. Banana *Musa acuminata*
2. Sugarcane *Saccharum officinarum*
3. Ginger *Zingiber officinale*
4. Medicinal plants *Chlorophytum borovillianum* (Safed musli), *Aloe barbadensis*
5. Ornamental plants *Carnation-Dianthus caryophyllus*, *Orchids-Vanilla*

**Location** The tissue culture laboratory should be preferably located in a moderated climate condition having uninterrupted supply of water and power. The tissue culture operations have to be carried out under controlled conditions of temperature. Extreme climatic condition adds to the cost of maintenance.

**Project Cost**

**A. Fixed asset**

**Table 2. Fixed asset of Tissue culture project**

Head	Cost ( Rs. In lack)
Land	5.00
Land development	5.60
Building	35.20
Utilities	16.00
Equipment	69.40
Green and shade house	30.00
Miscellaneous fixed asset	2.75
Total	163.95

**Land:**

Approximate 5 acres land should be adequate for setting up a TC unit with the above capacity. Cost of land is assumed at Rs. 5.00 Lakhs Building and civil works The building of about 8800 sq.ft includes class 1000 clean rooms and areas with comfort AC for laboratory, growth rooms and office space.

**The following facilities would be required in the building.**

- a) Storage room for chemicals
- b) Washing and Media preparation room
- c) Sterilization room
- d) Inoculation room
- e) Culture room

The total cost is estimated at Rs. 35.20 lakhs @ Rs. 400/sft. Green house A green house of 7500 sq.ft. and a shade house of 80,000 sq.ft. have been assumed at a cost of Rs. 22.00 lakhs and 8.00 lakhs (total Rs. 30 lakhs) respectively<sup>[15]</sup>.

**Equipment Major equipment and instruments required for the plant are as follows:**

- Autoclave
- Laminar air
- Flow cabinet
- Equipment for sterilization
- Electronic weighing balance
- Water distillation apparatus
- Air handling units
- Refrigerator
- Air conditioners
- Stereomicroscope
- Digital pH meter
- Shelves / racks
- Green house material

**WORKING CAPITAL REQUIREMENT**

**(I) Raw material**

The basic inputs for the production of micropropagated plantlets include meristems of elite and disease free plants, ready to use culture medium, sucrose and agar.

**(II) Manpower**

The unit with the proposed capacity may need 40-50 people at various positions including managerial, supervisory, skilled and unskilled<sup>[16]</sup>.

**Table 3. Recurring expenses (per month)**

(III) Recurring expenses (per month)	(Rs. lakhs)
Raw Material	2.50
Manpower	2.41
Utilities (power, water)	0.45
Contingencies (marketing, office expense, repair etc)	0.40
Total	5.76

**(B) ECONOMICS OF STARTING PLANT TISSUE CULTURE BUSINESS WITH THE MINIMAL INVESTMENT:**

Micro propagation business can be started by entrepreneurs interested in venturing into this area, with smaller investment by setting up a hardening unit to start with. Such entrepreneurs can procure primary hardened tissue culture plantlets from established micro propagation units and undertake secondary hardening in the facility and sell it to the farmers. Once the market is established, a full-fledged micro propagation unit could be set up. The following profile provides an overview of profitability for a hardening facility for handling 3 lakh plantlets per annum.

**4. Government Schemes and Incentives:** Various Central and State Government departments have framed financial schemes and announced incentives for assistance of tissue culture industry which are summarized below: a. Ministry of Agriculture The Department of Agriculture and Cooperation under the Ministry of Agriculture, Government of India

has the following programmes and schemes for promotion of horticulture<sup>[17]</sup>.

- (i) There is a provision for assistance of upto Rs. 21 lakhs and Rs. 10 lakh for setting up tissue culture units in public and private sector respectively subject to a maximum of 20% of the project cost.
- (ii) Under the Integrated Development of Fruits scheme assistance is given for purchase of planting material under the area expansion programme for the following crops: -
  - a) Rs. 7,000/hectare for plants of Guava, Amla, Date Palm, Plum Peach, Bes, Fig and citrus.
  - b) Rs.10,000/hectare for plants of mango, almond, pomegranate, apple, nuts, apricot, olive, papaya, litchi and sapota.
  - c) Rs. 30,000/hectare for plants for Bananas and pineapples.
  - d) Rs. 70,000/hectare for plants of grapes and strawberry.

In addition, 50% subsidy is given to the farmers for purchase of tissue culture banana by the Andhra Pradesh State Agriculture Department under the Macro Management Scheme

b. Agricultural and Processed food products Export Development Authority (APEDA) APEDA under the Ministry of Commerce and Industry has taken the following initiatives for promoting tissue culture in the country.

- (i) A state-of-the-art airfreight trans-shipment centre has been set up for temperature sensitive perishables at Delhi, Mumbai and Bangalore airports<sup>[18]</sup>.
- (ii) Airfreight subsidy is given for Tissue Culture Plants along with other live plants / bulb in category of perishable horticulture produce for export. The rate of subsidy to West Asia and CIS countries is at the rate of Rs.10 per kg or 25% of the airfreight rate approved by IATA or 1/3 rd of the FOB value whichever is the least.
- (iii) The rate of subsidy for export to Europe other than CIS countries, North America and Far East at the rate of Rs.25 per kg or 25% of the airfreight rate approved by IATA or 1/3 rd of the FOB value whichever is the least.
- (iv) 50% subsidy is given for the development of infrastructure like refrigerated van, packaging,

export promotion, market development, consultancy services and feasibility studies, organization building and human resource development.

(v) Financial assistance is also given for strengthening quality control facilities and implementation of ISO 9000.c. National Horticulture Board (NHB) The mandate of NHB is to promote integrated development of Horticulture and to help in coordinating, stimulating and sustaining the production and processing of fruits and vegetables. It also helps in establishing a sound infrastructure in the field of production, processing and marketing with a focus on post harvest management. For setting up of a new tissue culture lab there is a provision for back-ended capital subsidy not exceeding 20% of the project cost with a maximum limit of Rs. 25 lakh per project. NHB also has a scheme for providing subsidy for cultivation under controlled climate condition in poly houses, green houses, net houses, etc<sup>[19]</sup>. The units planning expansion in the domestic market by having a network of nurseries or additional hardening facilities can avail this scheme. The provision also exists for high quality commercial horticulture crops, Indigenous crops/produce, herbs, aromatic & medicinal plants, seed & nursery, bio-pesticide and establishment of Horticulture Health Clinics Laboratory. In all these cases, the subsidy is routed through the involvement of a financial institution on the completion of the project.<sup>[20]</sup>

## CONCLUSION

The current article combines the study of plant tissue culture performed by different scientists world wide. Plant tissue culture technique has brought revolution in the pharmacy field. This study unwinds different aspects of plant tissue culture technique and shows applicability of this tool for production of pharmaceuticals. It can be concluded that plants are the wide source of medicines. Tissue culture technique can be utilized for production of such medicines. Tissue culture started a new era in the field of phytochemicals. Though many techniques are developed till date to improve yield and economy of tissue culture, more research is should be carried out for further development.



**Fig II: Inoculation of excised micro shoots**

**Fig. 2. In vitro rooting of micro shoots.**



**Fig III: In vitro rooting of micro shoots**

**Fig. 3. Ex Agar plants.**



**Fig IV: Ex agar plants ready for packaging and dispatch**



**Fig. V: Hardening of plants in green house**

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